#### Amino - NHS ester lableing Protocol

This protocol is for labeling of oligonucleotides carrying an amine group with mono-NHS ester dyes.

This protocol is based on Rahul's and Jeehae's and modified by SH. Thanks.

The key concepts are

1) The optimal **amine:dye ratio is 1:5** (I prefer to use **1:20**)

2) Set pH of reaction buffer to 8.5 (High pH enhances the acylation rate and hydrolysis of the esters)

3) Typical amine concentration in reaction buffer is about 150uM. (but not critical)

## 1. Preperation

\* Recaction buffer (100mM Sodium tetraborate)

Dissolve 201 mg of sodium tetraborate anhydrous (or 380 mg for decahydrate) in 10ml dewater. (use fresh bufer) Adjust the pH to 8.5 with HCl. (usually need ~70ul of 12M HCl. Initially is at pH9.2)

\* Dye (final: 23mM in DMSO)

Dissolve 1mg of mono-NHS ester dye in 56 ul of DMSO. Dye in DMSO can be stored in -20 for 2 weeks.

\* Oligo (final concentration: 200uM in reaction buffer,100mM Na2B4O7 pH8.5)

ex. If you have 10 nMole of oligos, add 50ul of labeling buffer.

# 2. Reaction

Mix followings. The total amount can be varied but keep the ratio of contents. (NH3:dye=1:20)

- | 14ul of 22 mM dye in DMSO => 1.5 mM
- | 75 ul of 200 uM oligo carring on amine group => 150 uM
- | 11ul of diWater

## 3. Incubation

Incubate at RT for 6 hours on a gently shaking mixer in dark.

## 4. EtOH precipitation

a) Add 1/10 of reaction solution volume of 3M NaCl to reaction solution. Add 2.5 volume of reaction solution of cold (-20 chilled) absolute ethanol. Mix well and incubate in -20 for 30mins.

b) DNA precipitation results in truning the solution turbid. Centrifuge the solution at -~12,000g for 30mins to recover the DNA as a pellet.

c) Remove the supernatant and wash the pellet 3-4 times with 70% etOH.

#### 5. Redissolve the DNA according to your aplication.

#### 6. Labeling efficiency

Cy3 and Cy5 molar extinction coefficients are 150,000 and 250,000 M<sup>-1</sup>cm<sup>-1</sup>. Dye absorptions at 260nm are less than 10%.