

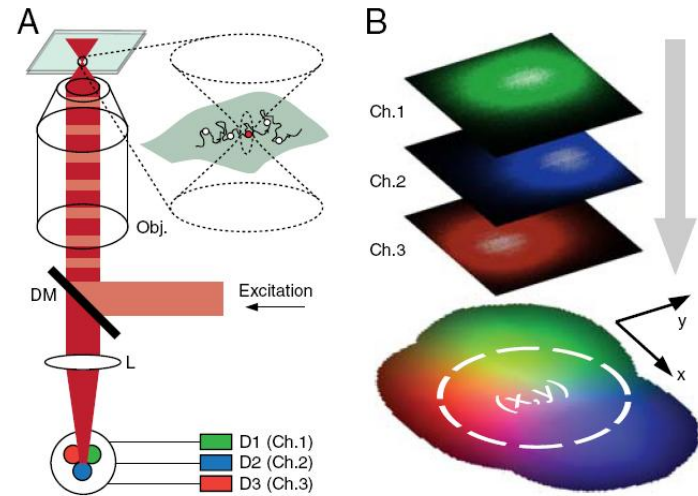
Fast molecular tracking maps nanoscale dynamics of plasma membrane lipids

Steffen J. Sahl¹, Marcel Leutenegger¹, Michael Hilbert, Stefan W. Hell², and Christian Eggeling²

Department of NanoBiophotonics, Max Planck Institute for Biophysical Chemistry, Am Fassberg 11, 37077 Göttingen, Germany

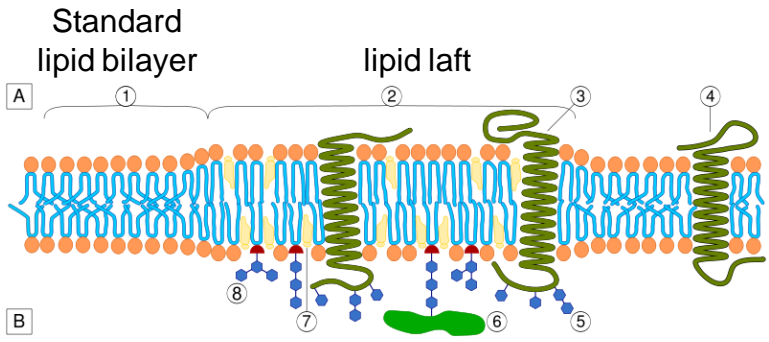
Edited by W. E. Moerner, Stanford University, Stanford, CA, and approved March 1, 2010 (received for review November 8, 2009)

We describe an optical method capable of tracking a single fluorescent molecule with a flexible choice of high spatial accuracy ($\sim 10\text{--}20$ nm standard deviation or $\sim 20\text{--}40$ nm full-width-at-half-maximum) and temporal resolution (<1 ms). The fluorescence signal during individual passages of fluorescent molecules through a spot of excitation light allows the sequential localization and thus spatio-temporal tracking of the molecule if its fluorescence is collected on at least three separate point detectors arranged in close proximity. We show two-dimensional trajectories of individual, small organic dye labeled lipids diffusing in the plasma membrane of living cells and directly observe transient events of trapping on <20 nm spatial scales. The trapping is cholesterol-assisted and much more pronounced for a sphingo- than for a phosphoglycero-lipid, with average trapping times of ~ 15 ms and <4 ms, respectively. The results support previous STED nanoscopy measurements and suggest that, at least for nontreated cells, the transient interaction of a single lipid is confined to macromolecular dimensions. Our experimental approach demonstrates that fast molecular movements can be tracked with minimal invasion, which can reveal new important details of cellular nano-organization.



Seoncheol Cha
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Journal Club

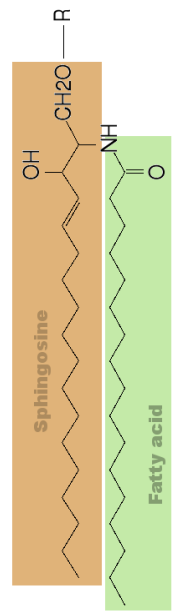
Lipid raft



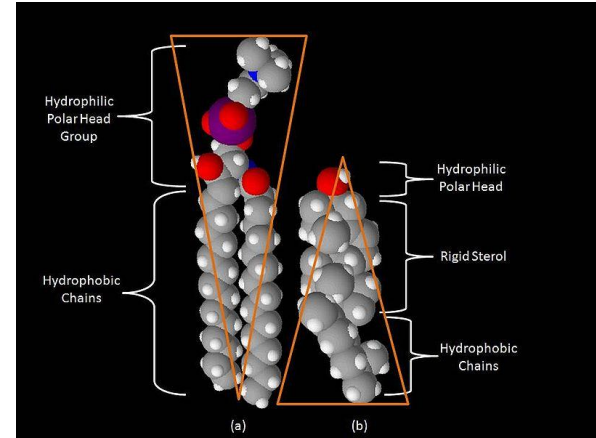
Lipid rafts are commonly defined as cholesterol- and sphingolipid-enriched membrane microdomains that function as platforms that concentrate and segregate proteins within the plane of the bilayer.

Studied by direct imaging(hard), smFRET, FRAP, single particle tracking, FCS ...

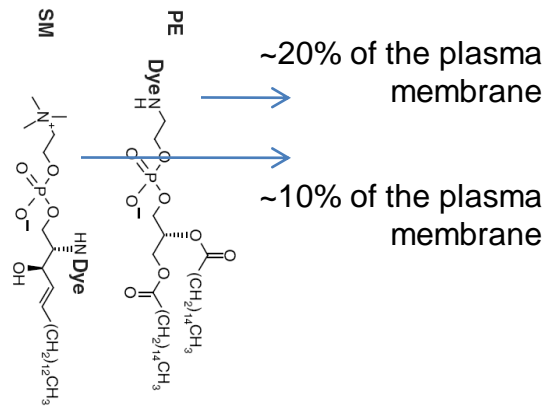
Neuropharmacology **55** 1265

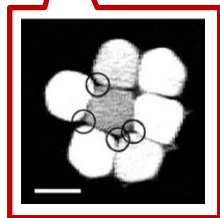
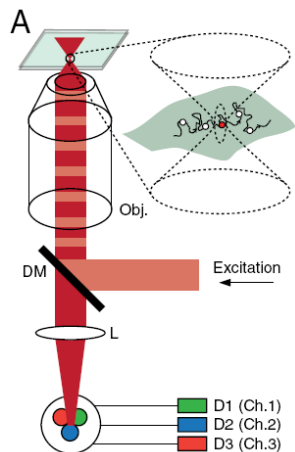


Wikipedia



Wikipedia

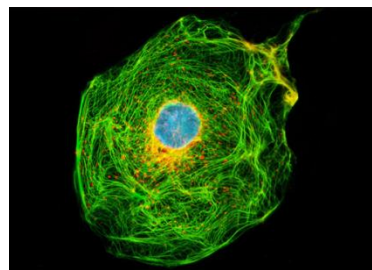
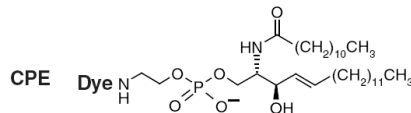
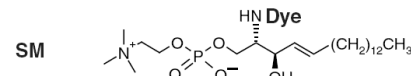
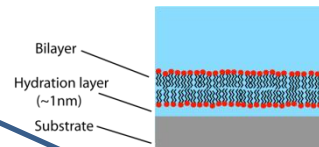
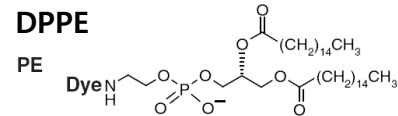
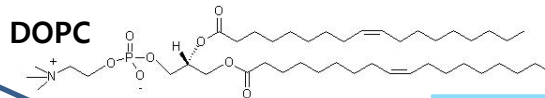




Scale bar :
125um

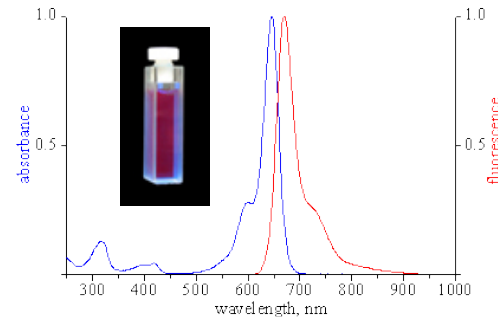
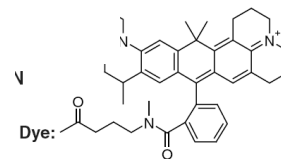
HeNe laser : 5~20uW 633nm (Melles Griot)
 100x NA=1.4 Oil immersion (Leica)
 APD (SPCM-AQR-13, Perkin elmer)
 TCSPC (SPC-830, Becker&Hickl)
 Router (HRT-82, Becker&Hickl)
 Fiber : multimode, 100um core diameter
 Closed-loop piezostage (for reference)

Supported Lipid bilayer

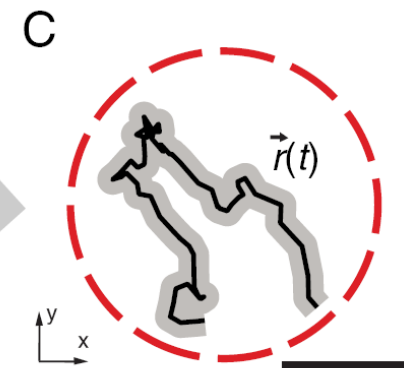
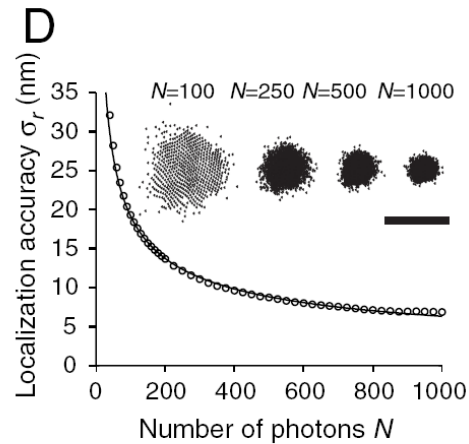
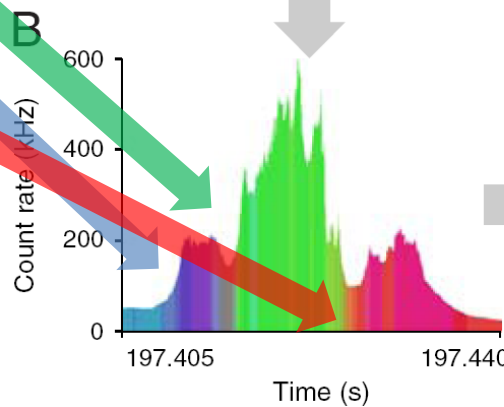
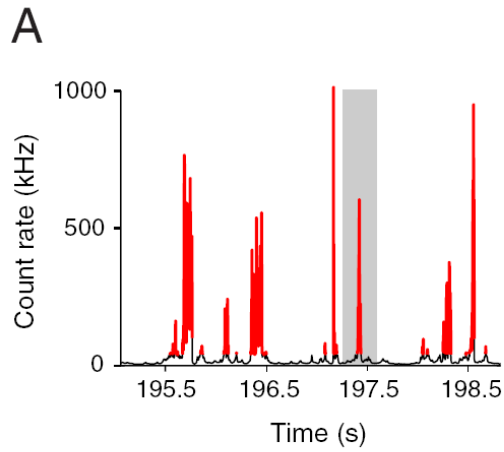
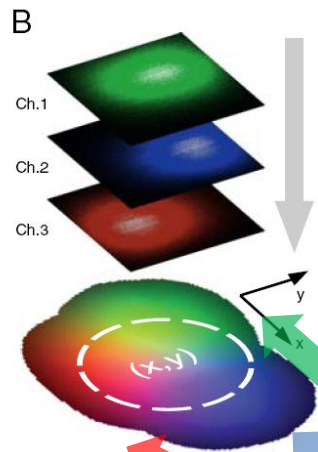
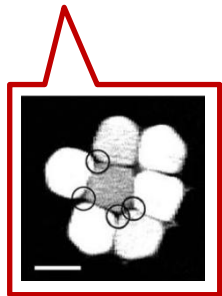
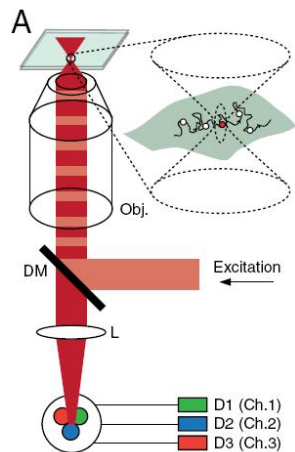


PtK2 Cell

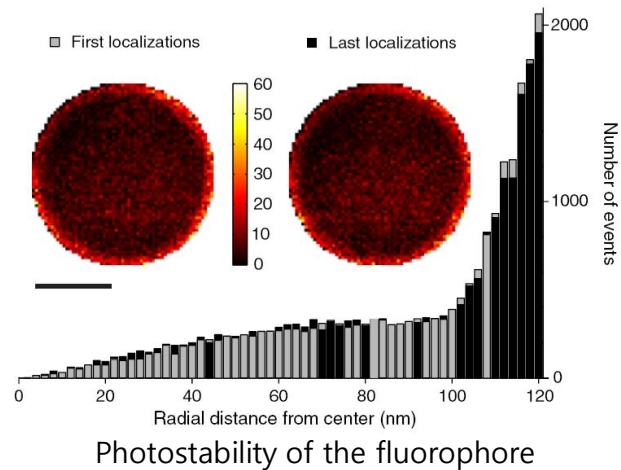
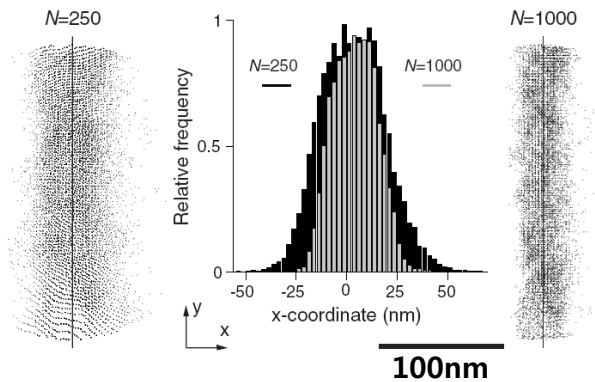
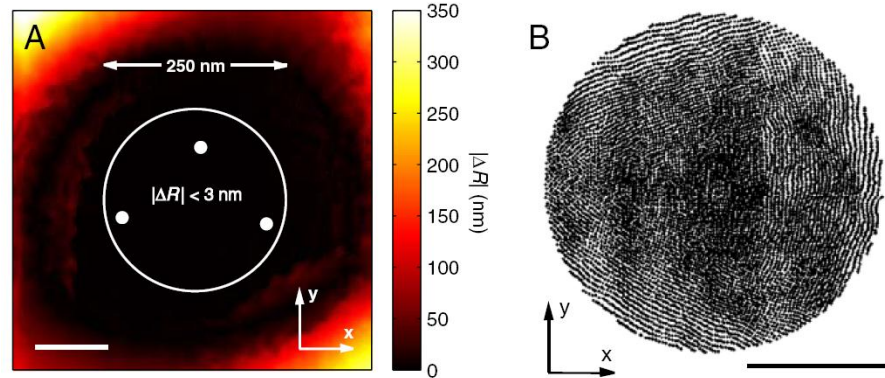
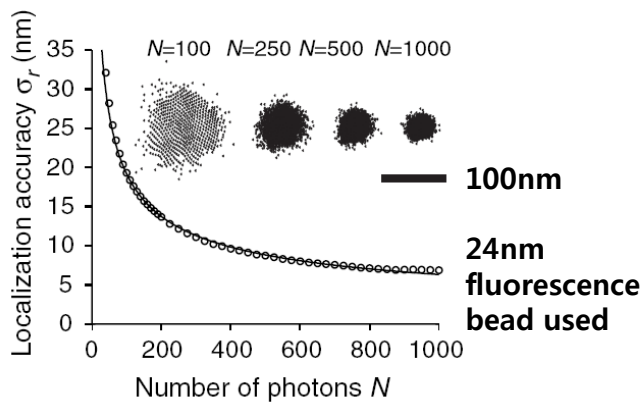
Incorporate

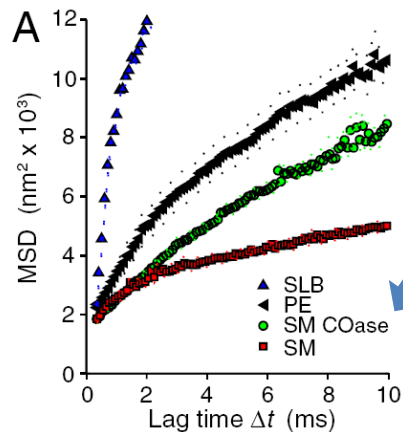


Direct Access to Biomolecular Dynamics by Fast Tracking



Localization Performance



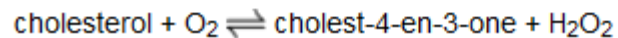


Mean Square Displacement

Cholesterol oxidase

From Wikipedia, the free encyclopedia

In enzymology, a **cholesterol oxidase** (EC 1.1.3.6 [🔗](#)) is an enzyme that catalyzes the chemical reaction



Analysis of Lipid Trajectory

Cumulant Probability

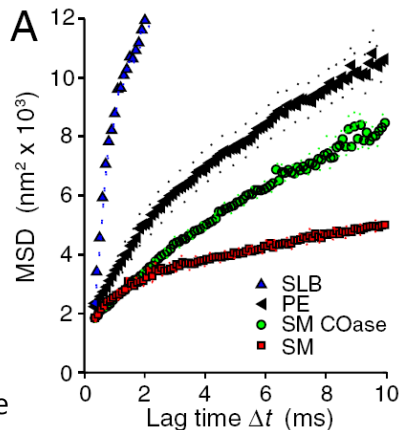
$P(\Delta r^2, \Delta t)$: probability that a molecule will be found within a circle of radius Δr from its starting point after time Δt

Not only to the **mean** but to the **full probability information**

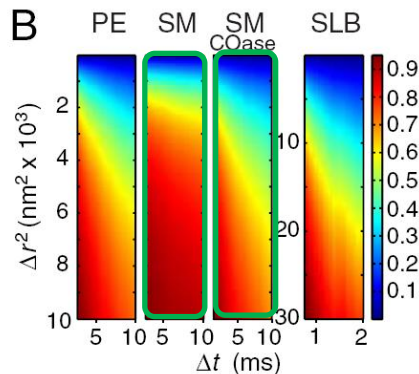
Directional flow
Lateral heterogeneities
Lateral constraints

Multi-component mobility

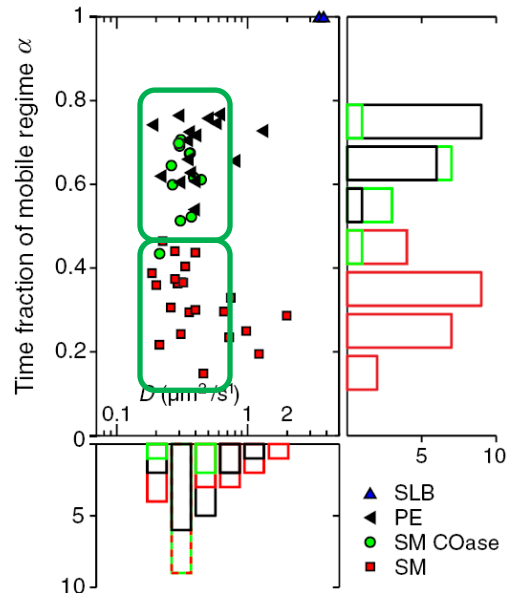
Biophys. J. **73** 1073



Mean Square Displacement



Cumulant Probability

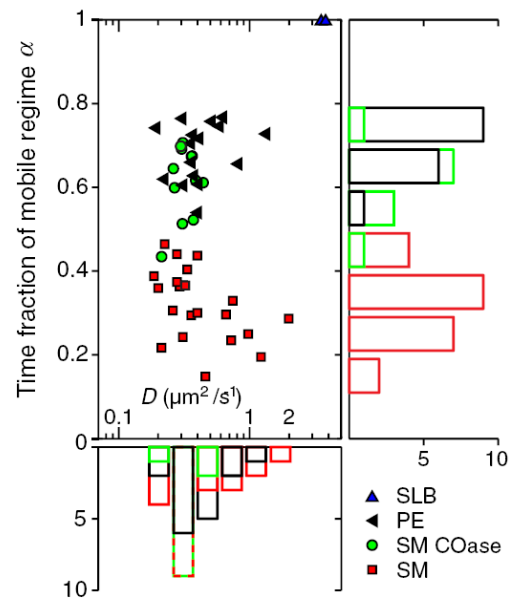
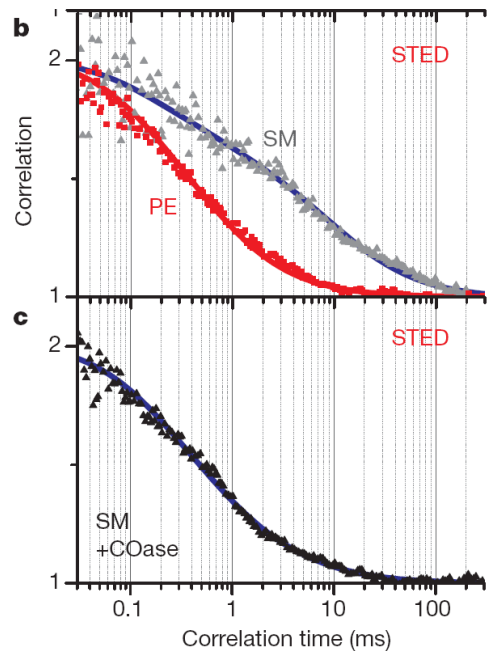


$$P(\Delta r^2, \Delta t) = \alpha \left[1 - \exp\left(\frac{-\Delta r^2}{4\sigma_r^2 + 4D\Delta t}\right) \right] + (1 - \alpha) \left[1 - \exp\left(\frac{-\Delta r^2}{4r_{\text{trap}}^2}\right) \right]$$

Free Brownian Motion

Hindered Motion

Results, and a comparison with STED-FCS



	$\alpha_{\text{STED-FCS}}$	$\alpha_{\text{fasttracking}}$
PE	1	0.7
SM	0.36	0.3
SM+COase	0.85	0.65

	D ($\mu\text{m}^2/\text{s}$)	α	r_{trap} (nm)	r_{trap}^* (nm)	\bar{t}_{trap} (ms)
SLB	3.6	1	-	-	-
PE	0.4	0.7	11	3	3
SM	0.35	0.3	6	3	17
SM+COase	0.3	0.65	9	3	3

Conclusion

High spatial localization accuracy (10-20nm)

Temporal resolution (0.5ms)

Applicable to lifetime, color, polarization anisotropy ...

Disadvantage is the limited observation area (~250nm)

-> limiting the length of the trajectories observed

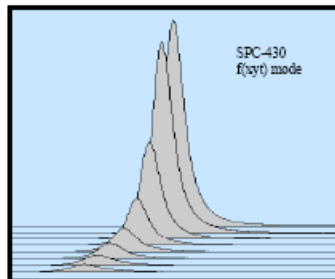
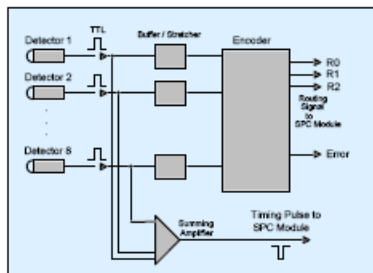
HRT-82

8 Channel TCSPC-Router for APD Modules

- Connects up to eight separate APD modules to one bh TCSPC module
- Simultaneous measurement in all detector channels
- Applicable with SPCM-AQR Modules and other TTL Output Detectors
- Count Rate > 3 MHz



The HRT-82 module is used to connect up to eight individual avalanche photodiode (APD) detectors to one of the time-correlated single photon counting modules SPC-xx0. The photons from the individual detectors are routed into different curves in the SPC memory. Thus the measurement yields a separate decay function for each of the detectors. Typical applications are fluorescence depolarisation measurements or simultaneous decay measurements at different wavelengths.

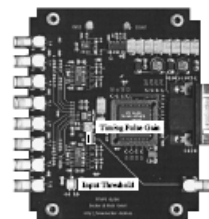


HRT-82

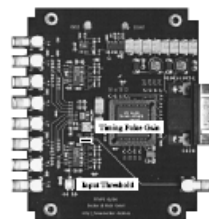
Specification

Input Polarity	positive
Input Voltage	TTL, 1.2 V to 5 V
Input Threshold	adjustable from 0.1 V to 2 V
Input Impedance	50 Ω
Input Pulse Duration	8 ns to 60 ns
Input Connectors	SMA
Timing Output Polarity	negative
Timing Output Voltage (2.5 V Input)	120 mV or 60 mV into 50 Ω (Jumper)
Timing Output Impedance	50 Ω
Timing Output Connector	50 Ohm, SMA
Delay Difference between Channels	max. 60 ps per Channel
Routing-Signal	TTL 3 bit + Error Signal
Routing Signal Connector	15 pin Sub-D/HD
Power Supply	+5V, -5V, via Sub-D Connector from SPC Module
Dimensions	120mm \times 95mm \times 34mm

Output Voltage Configuration

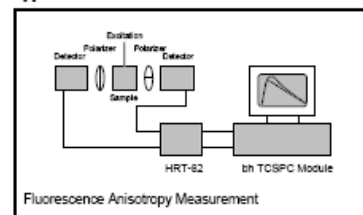


Vout = 120 ... 150 mV (SPC-xx30)

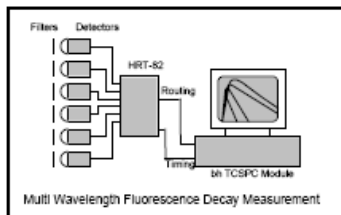


Vout = 50 ... 60 mV (SPC-xx0)

Applications



Fluorescence Anisotropy Measurement



Multi Wavelength Fluorescence Decay Measurement